

# **Triglycerides**

GPO-POD. Enzymatic colorimetric

# Quantitative determination of triglycerides

Store at 2-8°C

## PRINCIPLE OF THE METHOD

Sample triglycerides incubated with lipoproteinlipase (LPL), liberate glycerol and free fatty acids. Glycerolis converted to glycerol-3-phosphate (G3P) and adenosine-5-diphosphate (ADP) by glycerol kinase (GK) and ATP. Glycerol-3phosphate (G3P) is then converted by glycerol phosphate oxidase (GPO) to dihydroxyacetone phosphate (DAP) and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). In the last reaction, hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) reacts with 4-aminophenazone

(4-AP) and p-chlorophenol in presence of peroxidase (POD) to give a red colored dye

Triglycerides + 
$$H_2O$$
  $\xrightarrow{\text{LPL}}$  Glycerol + free fatty acids  
Glycerol + ATP  $\xrightarrow{\text{Glycerolkinase}}$  G3P + ADP  
G3P +  $O_2$   $\xrightarrow{\text{GPO}}$  DAP +  $H_2O_2$   
 $H_2O_2$  + 4-AP + p-Chlorophenol  $\xrightarrow{\text{POD}}$  Quinone +  $H_2O_2$ 

The intensity of the color formed is proportional to the triglycerides concentration in the sample <sup>1,2,3</sup>.

### CLINICAL SIGNIFICANCE

Triglycerides are fats that provide energy for the cell.

Like cholesterol, they are delivered to the body's cells by lipoproteins in the blood. A diet with a lot of saturated fats or carbohydrates will raise the triglyceride levels. The increases in serum triglycerides are relatively nonspecific. For example liver dysfunction resulting from hepatitis, extra hepatic biliary obstruction or cirrhosis, diabetes mellitus is associated with the increase

Clinical diagnosis should not be made on a single test result; it should integrate clinical and other laboratory data.

### REAGENTS

R1	GOOD pH 7,5	50 mmol/L
Buffer	p-Chlorophenol	2 mmol/L
	Lipoprotein lipase (LPL)	150000 U/L
	Glycerolkinase (GK)	500 U/L
R 2	Glycerol-3-oxidase (GPO)	2500 U/L
Enzymes (Note 2)	Peroxidase (POD)	440 U/L
	4 - Aminophenazone (4-AP)	0,1 mmol/L
	ATP	0,1 mmol/L
TRIGLYCERIDES CAL	Triglycerides aqueous primary standard 200 mg/dL	

## PREPARATION

Working reagent (WR): Dissolve ( $\rightarrow$ ) the contents of one vial R 2 Enzymes into one bottle of R 1 Buffer.

Cap and mix gently to dissolve contents.

WR stability: 6 weeks at 2-8°C or 1 week at room temperature (15-25°C).

# STORAGE AND STABILITY

All the components of the kit are stable until the expiration date on the label when stored tightly closed at 2-8°C, protected from light and contaminations prevented during their use. Do not use reagents over the expiration date

# Signs of reagent deterioration:

- Presence of particles and turbidity
- Blank absorbance (A) at 505 nm ≥ 0,14.

## ADDITIONAL EQUIPMENT

- Spectrophotometer or colorimeter measuring at 505 nm.
- Matched cuvettes 1,0 cm light path.
- General laboratory equipment.

## SAMPLES

Serum or heparinized or EDTA plasma1. Stability of the sample: 5 days at 2-8°C

# PROCEDURE

1.	Assay conditions:
	Wavelength:
	Cuvette:
	Temperature:
2.	Adjust the instrument to zero with distilled water.

- Pipette into a cuvette (Note 4):

	Blank	Standard	Sample
WR (mL)	1,0	1,0	1,0
Standard (Note 1,3) (µL)	(677.6)	10	750
Sample (µL)	100000	752	10

- Mix and incubate for 5 min. at 37°C or 10 min. at room temperature.
- Read the absorbance (A) of the samples and Standard, against the Blank. The colour is stable for at least 30 minutes.

### CALCULATIONS

(A) Sample - (A) Blank x 200 (Standard conc.) = mg/dL triglycerides in the sample (A) Standard - (A) Blank

Conversion factor: mg/dL x 0,0113= mmol/L.

### **QUALITY CONTROL**

Control sera are recommended to monitor the performance of assay procedures: SPINTROL H Normal and Pathologic (Ref. 1002120 and 1002210)

If control values are found outside the defined range, check the instrument, reagents and calibrator for problems.

Each laboratory should establish its own Quality Control scheme and corrective actions if controls do not meet the acceptable tolerances.

### REFERENCE VALUES

Men 40 - 160 mg/dL 35 - 135 mg/dL Women

These values are for orientation purpose; each laboratory should establish its own reference range.

### PERFORMANCE CHARACTERISTICS

Measuring range: From detection limit of 0,000 mg/dL to linearity limit of 1200

If the results obtained were greater than linearity limit, dilute the sample 1/2 with NaCl 9 g/L and multiply the result by 2.

#### Precision:

	Intra-ass	ay (n=20)
Mean (mg/dL)	103	219
SD	0,41	0,93
CV (%)	0,39	0,43

Inter-ass	ay (n=20)
103	217
3,74	7,80
3,62	3,59

Sensitivity: 1 mg/dL = 0,00137 A.

Accuracy: Results obtained using on SPINREACT reagents (y) did not show systematic differences when compared with other commercial reagents (x).

The results obtained using 50 samples were the following:

Correlation coefficient (r): 0,99760

Regression equation: y= 0,905x + 10,77.

The results of the performance characteristics depend on the analyzer used.

## INTERFERENCES

No interferences were observed with bilirubin up to 170  $\mu$ mol/L and hemoglobin up to 10 g/L2.

A list of drugs and other interfering substances with cholesterol determination has been reported by young etal 4.5.

### NOTES

- TRIGLYCERIDES CAL: Proceed carefully with this product because due its nature it can get contaminated easily.
- 2 LCF (Lipid Clearing Factor) is integrated in the reagent.
- 3. Calibration with the aqueous Standard may cause a systematic error in automatic procedures. In these cases, it is recommended to use a serum Calibrator.
- Use clean disposable pipette tips for its dispensation.
- 5. SPINREACT has instruction sheets for several automatic analyzers.

## **BIBLIOGRAPHY**

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- Kaplan A et al. Tryglycerides. Clin Chem The C.V. Mosby Co. St Louis. Toronto. Princeton 1984; 437 and Lipids 1194-1206. Young DS. Effects of drugs on Clinical Lab. Tests, 4th ed AACC Press, 1995.
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# **PACKAGING**

Ref: 1001310		R1: 1 x 50 mL, R2: 5 → 10 mL, CAL: 1 x 5 mL
Ref: 1001311		R1: 10 x 20 mL, R2: 10 → 20 mL, CAL: 1 x 5 mL
Ref: 1001312	Cont	R1: 10 x 50 mL, R2: 10 → 50 mL, CAL: 1 x 5 mL
Ref: 1001313	- 001110	R1: 4 x 125 mL, R2: 4 → 125 mL, CAL: 1 x 5 mL
Ref: 1001314		R1: 4 x 250 mL, R2: 4 → 250 mL, CAL: 1 x 5 mL